

## Sesquiterpenoids from the Resinous Exudates of *Commiphora opobalsamum* (Burseraceae)

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Phytochemical investigation of the exudates of *Commiphora opobalsamum* led to the isolation of three new sesquiterpenoids, 2 $\alpha$ -methoxy-6-oxogermacra-1(10),7(11)-dien-8,12-olide (**1**), 5 $\beta$ -10 $\alpha$ -hydroxy-2 $\alpha$ -methoxy-6-oxoguaia-7(11),8-dien-8,12-olide (**2**), and furanocadina-1(10),6,8-triene-4-ol (**3**), together with six known compounds. Their structures were elucidated on the basis of spectroscopic methods.

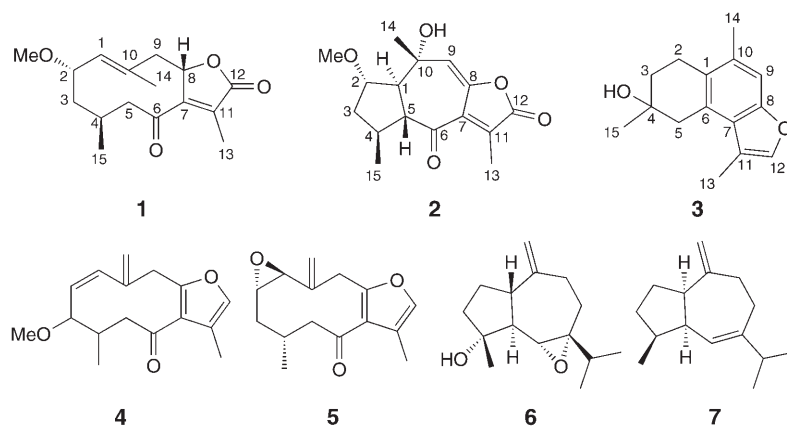
**Introduction.** – The genus *Commiphora*, belonging to the family Burseraceae, comprises over 150 species, and mainly occurs in Eastern Africa, Arabia and India [1]. The resinous exudates of these plant species possess diverse biological activities, such as cytotoxic, anaesthetic, anti-inflammatory, and antimicrobial effects, and have been used for a long time in the traditional medicines of India, China, Rome, Greece, and Babylon [2–10]. Previous phytochemical investigations of this genus resulted in the isolation of a series of terpenoids, steroids, flavonoids, and sugars [11][12].

In our continuous search for bioactive constituents from Chinese traditional medicinal plants [3], we herein report the three new sesquiterpenes **1–3**, along with six known compounds **4–9** from the resinous exudates of *C. opobalsamum*. Furthermore, the cytotoxic and antifungal activities of compounds **1–8** were evaluated against two human tumor cell lines and *Candida albicans*, respectively.

**Results and Discussion.** – The exudates of *C. opobalsamum* were extracted with petroleum ether (PE), and then purified by repeated column chromatography over silica gel and preparative TLC to afford three new sesquiterpenoids **1–3**, together with the known compounds **4–9**.

Compound **1** was isolated as colorless crystals, which showed a pseudomolecular ion at  $m/z$  301.6 [ $M + Na$ ]<sup>+</sup> in the positive-ion ESI-MS. Its molecular formula was determined to be C<sub>16</sub>H<sub>22</sub>O<sub>4</sub> by the  $M^+$  peak at  $m/z$  278.1527 (calc. 278.1518) in the HR-EI-MS. The <sup>1</sup>H-NMR spectrum showed the presence of two Me *singlets* ( $\delta$ (H) 1.75 and

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3.07), two Me doublets ( $\delta(\text{H})$  1.18 (*d*,  $J = 7.2$  Hz) and 2.01 (*d*,  $J = 1.8$  Hz)), two oxymethine H-atoms ( $\delta(\text{H})$  4.04 and 5.35–5.37), and one olefinic H-atom ( $\delta(\text{H})$  5.20 (*d*,  $J = 7.6$  Hz)). The  $^{13}\text{C}$ -NMR spectrum resolved 16 C-atom signals, which were determined by chemical shifts and the HMQC spectrum as four Me groups (one MeO group at  $\delta(\text{C})$  56.0), three  $\text{sp}^3$   $\text{CH}_2$ , and three  $\text{sp}^3$  CH groups (two oxygenated at  $\delta(\text{C})$  75.3 and 80.9), two CO groups ( $\delta(\text{C})$  197.9 and 172.8), and two C=C bonds (Table). The presence of an  $\alpha,\beta$ -unsaturated  $\alpha$ -methyl- $\gamma$ -lactone moiety was confirmed by the C-atom signals at  $\delta(\text{C})$  172.8 (C(12)), 129.6 (C(11)), 10.3 (C(13)), 158.7 (C(7)), and 80.9 (C(8)). The  $^1\text{H},^1\text{H}$ -COSY spectrum led to the identification of two partial structures:  $\text{CH}(1)\text{--CH}(2)\text{--CH}_2(3)\text{--CH}(4)\text{--Me}(15)$  and  $\text{CH}(8)\text{--CH}_2(9)$  (Fig. 1). Connection of these fragments to give a germacrane type lactone was rendered possible by the HMBC correlations of Me(14)/C(1), C(9), and C(10); H–C(5)/C(4) and C(6); Me(13)/C(6), C(7), C(11), and C(12); and H–C(8)/C(7) (Fig. 1). The position of the MeO group at C(2) was deduced from the cross-peak of H–C(2)/ $\delta(\text{C})$  56.0 (MeO).

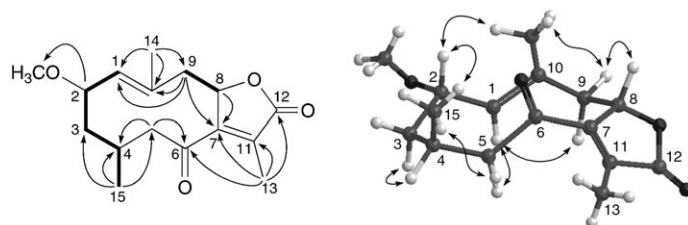


Fig. 1. Key  $^1\text{H},^1\text{H}$ -COSY (—), HMBC (H→C), and NOESY (H↔H) correlations of **1**

The relative configuration of **1** was established by a NOESY experiment (Fig. 1), in which the correlations of H–C(2)/Me(14) and Me(15);  $\text{H}_\beta\text{--C}(5)/\text{Me}(15)$ ;  $\text{H}_\beta\text{--C}(9)/\text{H--C}(8)$  and Me(14);  $\text{H}_\alpha\text{--C}(5)/\text{H--C}(1)$  and  $\text{H}_\alpha\text{--C}(9)$ ; and  $\text{H}_\alpha\text{--C}(3)/\text{H--C}(4)$  indicated that H–C(8), Me(15), and the MeO group at C(2) were  $\beta$ -,  $\beta$ -, and  $\alpha$ -

Table. <sup>1</sup>H- and <sup>13</sup>C-NMR Data of Compounds **1–3**. At 600/150 MHz, resp.; δ in ppm, *J* in Hz.

	<b>1<sup>a)</sup></b>		<b>2<sup>a)</sup></b>		<b>2<sup>b)</sup></b>		<b>3<sup>a)</sup></b>	
	δ(H)	δ(C)	δ(H)	δ(C)	δ(H)	δ(C)	δ(H)	δ(C)
H–C(1) or C(1)	5.20 ( <i>d</i> , <i>J</i> = 7.6)	133.6	2.31–2.38 ( <i>m</i> )	57.1	2.74 ( <i>br. d</i> , <i>J</i> = 6.3)	61.7	2.72–2.77 ( <i>m</i> ),	127.1
H–C(2) or CH <sub>2</sub> (2)	4.04 ( <i>dt</i> , <i>J</i> = 2.3, 7.4)	75.3	3.99–4.01 ( <i>m</i> )	82.1	4.19 ( <i>br. d</i> , <i>J</i> = 5.2)	83.7	2.83–2.89 ( <i>m</i> )	24.1
CH <sub>2</sub> (3)	1.88–1.93 ( <i>m</i> , H <sub>a</sub> ), 1.77–1.79 ( <i>m</i> , H <sub>β</sub> )	35.7	1.99–2.03 ( <i>m</i> , H <sub>a</sub> ), 1.41–1.46 ( <i>m</i> , H <sub>β</sub> )	38.9	2.12 ( <i>ddd</i> , <i>J</i> = 5.7, 13.3, H <sub>a</sub> ), 1.26–1.31 ( <i>m</i> , H <sub>β</sub> )	41.5	1.83–1.88 ( <i>m</i> ), 1.95–2.00 ( <i>m</i> )	35.3
H–C(4) or C(4)	2.28–2.32 ( <i>m</i> )	25.0	2.83–2.85 ( <i>m</i> )	31.3	2.95–3.00 ( <i>m</i> )	34.1		68.3
CH <sub>2</sub> (5) or H–C(5)	2.80–2.83 ( <i>m</i> , H <sub>a</sub> ), 2.53 ( <i>ddd</i> , <i>J</i> = 6.3, 17.5, H <sub>β</sub> )	47.2	2.31–2.38 ( <i>m</i> )	60.2	2.56 ( <i>t</i> , <i>J</i> = 10.2)	63.5	3.22–3.29 ( <i>m</i> )	41.0
C(6)		197.9		193.5		195.8		127.8
C(7)		158.7		144.8		145.7		124.6
H–C(8) or C(8)	5.35–5.37 ( <i>m</i> )	80.9		139.1		140.7		154.0
CH <sub>2</sub> (9) or H–C(9)	2.10 ( <i>t</i> , <i>J</i> = 11.8, H <sub>a</sub> ), 2.80–2.83 ( <i>m</i> , H <sub>β</sub> )	45.8	5.97 ( <i>s</i> )	121.2	6.27 ( <i>s</i> )	124.5	7.13 ( <i>s</i> )	110.3
C(10)		133.6		72.6		72.5		133.0
C(11)		129.6		137.9		137.5		115.7
C(12) or H–C(12)		172.8		168.9		170.2	7.26 ( <i>s</i> )	140.6
Me(13)	2.01 ( <i>d</i> , <i>J</i> = 1.8)	10.3	2.35 ( <i>s</i> )	11.0	2.20 ( <i>s</i> )	11.7	2.38 ( <i>s</i> )	11.0
Me(14)	1.75 ( <i>s</i> )	17.4	1.42 ( <i>s</i> )	22.3	1.53 ( <i>s</i> )	23.6	2.32 ( <i>s</i> )	20.2
Me(15)	1.18 ( <i>d</i> , <i>J</i> = 7.2)	22.8	1.09 ( <i>d</i> , <i>J</i> = 6.7)	19.7	1.16 ( <i>d</i> , <i>J</i> = 6.5)	19.6	1.41 ( <i>s</i> )	28.4
MeO	3.07 ( <i>s</i> )	56.0	3.32 ( <i>s</i> )	55.5	3.33 ( <i>s</i> )	56.9		

<sup>a)</sup> In CDCl<sub>3</sub>. <sup>b)</sup> In (D<sub>5</sub>)pyridine.

oriented, respectively. Therefore, the structure of **1** was assigned to be 2 $\alpha$ -methoxy-6-oxogermacra-1(10),7(11)-dien-8,12-olide<sup>2)</sup>, but its absolute configuration remains to be established.

Compound **2**, a white amorphous powder, exhibited a molecular ion peak  $M^+$  at  $m/z$  292.1310 (calc. 292.1311) in the HR-EI-MS, which is consistent with the molecular formula  $C_{16}H_{20}O_5$ . The  $^1H$ -NMR spectrum showed the presence of three Me *singlets* ( $\delta(H)$  1.42, 2.35, and 3.32), one Me *doublet* ( $\delta(H)$  1.09,  $d, J = 6.7$  Hz), and one olefinic H-atom ( $\delta(H)$  5.97, *s*). The  $^{13}C$ -NMR spectrum displayed signals for 16 C-atoms. By HMQC, they were determined to consist of four Me groups (one MeO group at  $\delta(C)$  55.5), one  $sp^3$   $CH_2$  ( $\delta(C)$  38.9), and four  $sp^3$  CH groups (one oxygenated at  $\delta(C)$  82.1), one oxygenated quaternary  $sp^3$  C-atom ( $\delta(C)$  72.6), two CO groups ( $\delta(C)$  193.5 and 168.9), one trisubstituted C=C bond, and one tetrasubstituted C=C bond (*Table*). The C-atom signals at  $\delta(C)$  168.9 (C(12)), 137.9 (C(11)), 11.0 (C(13)), 144.8 (C(7)), and 139.1 (C(8)), were undoubtedly assigned to an  $\alpha,\beta$ -unsaturated  $\alpha$ -methyl- $\gamma$ -lactone ring. The  $^1H,^1H$ -COSY spectrum of **2** confirmed the presence of a five-membered ring (depicted with bold lines in *Fig. 2*). Combined with the HMBC correlations of Me(14)/C(1), C(9), and C(10); H–C(9)/C(7) and C(8); H–C(5)/C(1) and C(6); and Me(13)/C(7), C(11), and C(12) (*Fig. 2*), compound **2** was established as a guaiane type lactone.

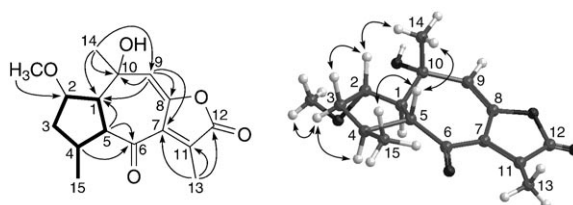


Fig. 2. Key  $^1H,^1H$ -COSY (—), HMBC (H  $\rightarrow$  C), and NOESY (H  $\leftrightarrow$  H) correlations of **2**

Due to signal overlapping of H–C(1) with H–C(5) at  $\delta(H)$  2.31–2.38 (*m*) in  $CDCl_3$ , we recorded the  $^1H$ -NMR spectrum in  $(D_5)$ pyridine, which caused that the H–C(1) resonance was shifted downfield *ca.*  $\Delta\delta$  0.39 to  $\delta(H)$  2.74, while H–C(5) was shifted downfield  $\Delta\delta$  0.21 to  $\delta(H)$  2.56 (*Table*). The relative configuration of **2** was established by a NOESY experiment in  $(D_5)$ pyridine. The NOESY correlations (*Fig. 2*) of H–C(2)/ $H_\beta$ -C(3) and Me(14);  $H_\alpha$ -C(3)/H–C(4) and the MeO group at C(2); and H–C(5)/Me(14) and Me(15) showed that H–C(2), H–C(5), Me(14), and Me(15) were  $\beta$ -oriented, whereas H–C(1), the OH group at C(10), and the MeO group at C(2) were  $\alpha$ -oriented. Thus, the structure of **2** was deduced as 5 $\beta$ -10 $\alpha$ -hydroxy-2 $\alpha$ -methoxy-6-oxoguaia-7(11),8-dien-8,12-olide<sup>2)</sup>.

Compound **3** was isolated as a yellow oil. The molecular formula was determined to be  $C_{15}H_{18}O_2$  on the basis of the  $M^+$  peak at  $m/z$  230.1299 (calc. 230.1307) in the HR-EI-MS. The  $^1H$ -NMR spectrum showed the presence of three Me *singlets* ( $\delta(H)$  1.41, 2.32, and 2.38), and two downfield signals at  $\delta(H)$  7.26 (1 H, *s*) and 7.13 (1 H, *s*). The  $^{13}C$ -NMR spectrum exhibited signals for 15 C-atoms including three Me groups ( $\delta(C)$

<sup>2)</sup> For systematic names, see *Exper. Part*.

11.0, 20.2, and 28.4), three CH<sub>2</sub> groups ( $\delta(C)$  24.1, 35.3, and 41.0), and one oxygenated quaternary C-atom ( $\delta(C)$  68.3), two trisubstituted C=C bonds, and two tetrasubstituted C=C bonds (Table). The <sup>1</sup>H- and <sup>13</sup>C-NMR data suggested that the structure of **3** was very similar to that of the known furanosesquiterpenoid dihydropyrocuzerenone [13], which was verified by analysis of its HMQC, <sup>1</sup>H,<sup>1</sup>H-COSY, and HMBC spectra (Fig. 3). The only difference was that the resonance for C(4) was shifted from  $\delta(C)$  28.6 to  $\delta(C)$  68.3, indicating that the methine H-atom at C(4) in dihydropyrocuzerenone was replaced by an OH group in **3**. Therefore, the structure of **3** was established as furanocadina-1(10),6,8-triene-4-ol<sup>2</sup>).

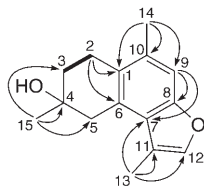


Fig. 3. Key <sup>1</sup>H,<sup>1</sup>H-COSY (—) and HMBC (H → C) correlations of **3**

The known compounds **4–9** were identified as (1*E*)-3-methoxy-8,12-epoxygermacra-1,7,10,11-tetraen-6-one (**4**) [13], *rel*-(1*S*,2*S*)-epoxy-(4*R*)-furanogermacr-10(15)-en-6-one (**5**) [14], 6*α*,7*α*-epoxy-1*β*-guaia-10(14)-en-4*α*-ol (**6**) [15], (1*R*,4*S*,5*R*)-guaia-6,10(14)-diene (**7**) [16], cerotic acid (**8**) [17], and  $\beta$ -sitosterol (**9**) [17] by comparison of their <sup>1</sup>H- and <sup>13</sup>C-NMR data with those reported in the cited references.

The antifungal activities of compounds **1–8** were tested against *Candida albicans*. Unfortunately, all test compounds were inactive (MIC > 64  $\mu$ g/ml). The cytotoxicities of **1–8** were evaluated against the PC3 (human prostate tumor) and HT29 (human colorectal carcinoma) cell lines. However, they were inactive against both human tumor cell lines (*IC*<sub>50</sub> > 50  $\mu$ M).

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### Experimental Part

**General.** Column chromatography (CC): silica gel (200–300 mesh, 10–40  $\mu$ m; Qingdao Haiyang Chemical Co. Ltd., China). TLC: silica gel GF<sub>254</sub> plates; visualization by heating the plates sprayed with 10% H<sub>2</sub>SO<sub>4</sub>/EtOH. M.p.: X-6 melting-point apparatus (Beijing TECH Instrument Co. Ltd., China). Optical rotations: Perkin-Elmer 241 MC polarimeter. UV: Agilent 8453E UV/VIS Spectrometer. IR: Nexus 470 FT-IR Spectrometer. 1D- and 2D-NMR spectra: Bruker Avance 600 spectrometer at 600 (<sup>1</sup>H) and 150 (<sup>13</sup>C) MHz. HR-EI-MS spectra: Waters GCT mass spectrometer. ESI-MS spectra: API 4000 mass spectrometer.

**Plant Material.** The exudates of *C. opobalsamum* were purchased in September 2002 from Affiliated Hospital of Shandong Traditional Chinese Medical University, Jinan, P. R. China. It was imported from India and identified by Prof. Qi-Shi Sun, School of Traditional Chinese Materia Medica, Shenyang Pharmaceutical University, P. R. China. A voucher specimen (No. 20020910CO) has been deposited at the Laboratory of Natural Products Chemistry, School of Pharmaceutical Sciences, Shandong University, P. R. China.

**Extraction and Isolation.** The resinous exudates of *C. opobalsamum* (3.5 kg) were extracted with petroleum ether (PE) in a Soxhlet apparatus for 36 h. The crude extract (290 g) was subjected to CC (SiO<sub>2</sub>; PE/AcOEt 1:0 → 0:1) to provide nine fractions (*Fr. A–I*). *Fr. B* (25 g) was subjected to CC (SiO<sub>2</sub>; PE/AcOEt 98:2 → 1:1) to afford nine subfractions (*Fr. B1–B9*). *Fr. B4* was separated by CC (SiO<sub>2</sub>; PE/Et<sub>2</sub>O 95:5 → 1:1) to give **8** (30 mg) and other four subfractions (*Fr. B4a–B4d*). **5** (15 mg) was isolated from *Fr. B4d* by preparative TLC (SiO<sub>2</sub>; PE/AcOEt 3:2). **4** (45 mg) was obtained from *Fr. B4a* by preparative TLC (SiO<sub>2</sub>; PE/CHCl<sub>3</sub>/Et<sub>2</sub>O 6:4:1). **7** (23 mg) was isolated from *Fr. B4c* by preparative TLC (SiO<sub>2</sub>; PE/AcOEt 13:7). *Fr. B8* was subjected to CC (SiO<sub>2</sub>; PE/AcOEt 1:1) to afford **9** (80 mg). *Fr. C* (21 g) was chromatographed over CC (SiO<sub>2</sub>; PE/AcOEt 97:3 → 1:1) to give five subfractions (*Fr. C1–C5*). **3** (5 mg) was obtained from *Fr. C5* by CC (SiO<sub>2</sub>; PE/Et<sub>2</sub>O 97:3), and purified by prep. TLC (SiO<sub>2</sub>; PE/CHCl<sub>3</sub>/Et<sub>2</sub>O 10:7:3). *Fr. D* was subjected to CC (silica gel; PE/Me<sub>2</sub>O 97:3 → 1:1) to give seven subfractions (*Fr. D1–D7*). *Fr. D6* was separated by CC (SiO<sub>2</sub>; PE/AcOEt 4:1) to afford **6** (8 mg). **1** (17 mg) was isolated from *Fr. D7* by CC (SiO<sub>2</sub>; PE/Me<sub>2</sub>O 9:1), and purified by recrystallization (PE/Me<sub>2</sub>O). Compound **2** (6 mg) was obtained from *Fr. E* by CC (SiO<sub>2</sub>; PE/Me<sub>2</sub>O 93:7), and then purified by CC (SiO<sub>2</sub>; PE/Et<sub>2</sub>O/Me<sub>2</sub>O 50:50:1).

**2 $\alpha$ -Methoxy-6-oxogermacra-1(10),7(11)-dien-8,12-olide** (= (6S,8S,11aS)-5,6,7,8,11,11a-Hexahydro-8-methoxy-3,6,10-trimethylcyclodeca[b]furan-2,4-dione; **1**). Colorless crystals (PE/Me<sub>2</sub>O). [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +196.2 (*c* = 0.053, CHCl<sub>3</sub>). UV (MeOH): 229.0 (4.08). IR (KBr): 2946, 2924, 2906, 2882, 1755, 1676, 1366, 1171, 1093, 1023, 753. <sup>1</sup>H- and <sup>13</sup>C-NMR: Table. ESI-MS: 301.6 (6, [M + Na]<sup>+</sup>), 296.5 (100, [M + NH<sub>4</sub>]<sup>+</sup>), 279.5 (40, [M + H]<sup>+</sup>). HR-EI-MS: 278.1527 (*M*<sup>+</sup>, C<sub>16</sub>H<sub>22</sub>O<sub>4</sub><sup>+</sup>; calc. 278.1518).

**5 $\beta$ -10 $\alpha$ -Hydroxy-2 $\alpha$ -methoxy-6-oxoguaia-7(11),8-dien-8,12-olide** (= (4aR,5S,7S,7aS,8S)-4a,5,6,7,7a,8-Hexahydro-8-hydroxy-7-methoxy-3,5,8-trimethylazuleno[6,5-b]furan-2,4-dione; **2**). White amorphous powder. [ $\alpha$ ]<sub>D</sub><sup>20</sup> = -46.1 (*c* = 0.065, CHCl<sub>3</sub>). UV (MeOH): 220.0 (3.82), 304 (3.89). IR (KBr): 3392, 3025, 2960, 2871, 1765, 1731, 1643, 1377, 767. <sup>1</sup>H- and <sup>13</sup>C-NMR: Table. ESI-MS: 315.4 (58, [M + Na]<sup>+</sup>), 310.7 (100, [M + NH<sub>4</sub>]<sup>+</sup>), 275.6 (75, [M + H - H<sub>2</sub>O]<sup>+</sup>). HR-EI-MS: 292.1310 (*M*<sup>+</sup>, C<sub>16</sub>H<sub>20</sub>O<sub>5</sub><sup>+</sup>; calc. 292.1311).

**Furanocadina-1(10),6,8-triene-4-ol** (= (6,7,8,9-Tetrahydro-1,5,8-trimethylnaphtho[2,1-b]furan-8-ol; **3**). Yellow oil. [ $\alpha$ ]<sub>D</sub><sup>20</sup> = -27.2 (*c* = 0.058, CHCl<sub>3</sub>). UV (MeOH): 215.0 (4.21), 246.0 (3.80), 253 (3.79). IR (KBr): 3428, 2951, 2924, 2866, 1615, 1572, 1538, 1453, 1101, 1029, 842, 766. <sup>1</sup>H- and <sup>13</sup>C-NMR: Table. HR-EI-MS: 230.1299 (*M*<sup>+</sup>, C<sub>15</sub>H<sub>18</sub>O<sub>2</sub><sup>+</sup>; calc. 230.1307).

## REFERENCES

- [1] K. Vollesen, in 'Burseraceae, Flora of Ethiopia', Addis Ababa University Press, Addis Ababa, 1989, Vol. 3, p. 442.
- [2] S. Habtemariam, *Toxicon* **2003**, *41*, 723.
- [3] T. Shen, W. Wan, H. Yuan, F. Kong, H. Guo, P. Fan, H. Lou, *Phytochemistry* **2007**, *68*, 1331.
- [4] M. Shoemaker, B. Hamilton, S. H. Dairkee, I. Cohen, M. J. Campbell, *Phytother. Res.* **2005**, *19*, 649.
- [5] P. Dolara, C. Luceri, C. Ghelardini, C. Monserrat, S. Aiolfi, F. Luceri, M. Lodovici, S. Menichetti, M. N. Romanelli, *Nature* **1996**, *379*, 29.
- [6] M. R. Meselhy, *Phytochemistry* **2003**, *62*, 213.
- [7] J. A. Francis, S. N. Raja, M. G. Nair, *Chem. Biodivers.* **2004**, *1*, 1842.
- [8] H. Matsuda, T. Morikawa, S. Ando, H. Oominami, T. Murakami, I. Kimura, M. Yoshikawa, *Bioorg. Med. Chem.* **2004**, *12*, 3037.
- [9] M. A. Saeed, A. W. Sabir, *Fitoterapia* **2004**, *75*, 204.
- [10] P. Dolara, B. Corte, C. Ghelardini, A. M. Pugliese, E. Cerbai, S. Menichetti, A. Lo Nostro, *Planta Med.* **2000**, *66*, 356.
- [11] E. S. H. El Ashry, N. Rashed, O. M. Salama, A. Saleh, *Pharmazie* **2003**, *58*, 163.
- [12] L. O. Hanuš, T. Režanka, V. M. Dembitsky, A. Moussaieff, *Biomed. Papers* **2005**, *149*, 3.
- [13] A. Dekebo, E. Dagne, O. Sterner, *Fitoterapia* **2002**, *73*, 48.
- [14] N. Zhu, H. Kikuzaki, S. Sheng, S. Sang, M. M. Rafi, M. Wang, N. Nakatani, R. S. DiPaola, R. T. Rosen, C.-T. Ho, *J. Nat. Prod.* **2001**, *64*, 1460.

- [15] M. Yoshikawa, S. Hatakeyama, N. Tanaka, Y. Fukuda, N. Murakami, J. Yamahara, *Chem. Pharm. Bull.* **1992**, *40*, 2582.
- [16] M. R. Rao, K. V. Sridevi, U. Venkatesham, T. P. Rao, S. S. Lee, Y. Venkateswarlu, *J. Chem. Res.* **2000**, 245.
- [17] Z. Ma, L. Qiu, *Acta Bot. Sin.* **1995**, *37*, 574.

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